

A close-up photograph of a fish's head, likely a cichlid, showing a large, textured, reddish-brown nuptial hump on its forehead. The fish's scales are iridescent, with shades of blue, purple, and red. The background is a solid blue color.

# International Journal of Fisheries and Aquaculture

Volume 7 Number 3 March 2015

ISSN 2006-9839



*Academic  
Journals*

## ABOUT IJFA

The **International Journal of Fisheries and Aquaculture (IJFA)** (ISSN: 2006-9839) is an open access journal that provides rapid publication (monthly) of articles in all areas of the subject such as algaculture, Mariculture, fishery in terms of ecosystem health, Fisheries acoustics etc.

The Journal welcomes the submission of manuscripts that meet the general criteria of significance and scientific excellence. Papers will be published shortly after acceptance. All articles published in the IJFA are peer-reviewed.

### Contact Us

**Editorial Office:** [ijfa@academicjournals.org](mailto:ijfa@academicjournals.org)

**Help Desk:** [helpdesk@academicjournals.org](mailto:helpdesk@academicjournals.org)

**Website:** <http://www.academicjournals.org/journal/IJFA>

**Submit manuscript online** <http://ms.academicjournals.me/>

## Editors

**Prof. Ashraf Mohamed Abd El-Samee Goda**  
*Fish Nutrition Research Laboratory,  
National Institute of Oceanography and  
Fisheries (NIOF), Cairo,  
Egypt.*

**Prof. Upali S. Amarasinghe**  
*Department of Zoology,  
University of Kelaniya,  
Kelaniya 11600, Sri Lanka.  
Sri Lanka.*

**Dr. V.S. Chandrasekaran**  
*Central Institute of Brackishwater Aquaculture  
(ICAR)  
75, Santhome High Road, R.A.Puram  
Chennai-600028,  
India.*

**Prof. Nihar Rajan Chattopadhyay**  
*Department of aquaculture,  
Faculty of Fishery Sciences,  
West Bengal University of Animal & Fishery  
Sciences,  
5. Buderhat Road,  
P.O. Panchasayar, Kolkata 700094,  
West Bengal,  
India.*

**Dr. Lourdes Jimenez-Badillo**  
*Ecology and Fisheries Centre, General  
Direction of Investigation,  
Universidad Veracruzana,  
Hidalgo 617, Col. Río Jamapa,  
Boca del Río,  
Veracruz, México ZP 94290.*

**Dr. Kostas Kapiris**  
*Institute of Marine Biological  
Resources of H.C.M.R.,  
Athens, Hellas  
Greece.*

**Dr. Masoud Hedayatifard**  
*Department of Fisheries Sciences and Aquaculture  
College of Agriculture and Natural Resources  
Advanced Education Center  
Islamic Azad University, Ghaemshahr,  
PO Box: 163,  
Iran.*

**Dr. Zhang Xiaoshuan**  
*209#, China Agricultural University(East campus),  
No.17 Qinghua Donglu,  
Beijing, China*

**Dr Joseph Selvin**  
*Marine Bioprospecting Lab  
Dept of Microbiology  
Bharathidasan University  
Tiruchirappalli 620 024  
India.*

## Editorial Board

**Dr. Dada Adekunle Ayokanmi**  
*Department of Fisheries and Aquaculture  
Technology, Federal University of  
Technology, P.M.B 704,  
Akure, Ondo State,  
Nigeria.*

**Dr. Ramasamy Harikrishnan**  
*KOSEF Post Doctoral Fellow, Faculty of  
Marine Science, College of Ocean Sciences,  
Jeju National University,  
Jeju city, Jeju 690 756,  
South Korea.*

**Dr. Kawser Ahmed**  
*Lab. of Ecology, Environment and  
Climate Change, Department of Fisheries,  
University of Dhaka, Dhaka-1000,  
Bangladesh.*

**Dr. Maxwell Barson**  
*Biological Sciences Department  
University of Zimbabwe PO Box MP 167  
Mount Pleasant Harare,  
Zimbabwe.*

**Dr. Christopher Marlowe Arandela Caipang**  
*Faculty of Biosciences and Aquaculture, Bodø University  
College, Bodø 8049,  
Norway.*

**Dr. William S. Davidson**  
*Department of Molecular Biology and Biochemistry  
Simon Fraser University 8888 University Drive  
Burnaby, British Columbia  
Canada V5A 1S6.*

**Dr. Babak Ghaednia**  
*Iran Shrimp Research Center ( ISRC)  
Taleghani High Way, P.O.Box 1374  
Bushehr,  
Iran.*

**Dr. Ramachandra Bhatta**  
*Animal and Fisheries Sciences University,  
College of Fisheries,  
Kankanady Mangalore 575 002  
India.*

**Dr. Harikrishnan**  
*Faculty of Marine Science  
College of Ocean Sciences  
Jeju National University, Jeju, 690-756  
South Korea .*

**Prof. Ratha Braja Kishore**  
*Department of Zoology  
Biochemical Adaptation Laboratory  
Banaras Hindu University  
Varanasi 221005  
India.*

**Dr. Esmail AM Shakman**  
*Am Vögenteich,13/ 3.09.618057 Rostock  
Germany .*

**Prof B. Sharma**  
*Department of Biochemistry  
Coordinator, Centre for Biotechnology  
University of Allahabad  
Allahabad-U.P.,  
India.*

**Dr. Sebastián Villasante**  
*Fisheries Economics and Natural Resources Research Unit  
University of Santiago de Compostela, A Coruña.  
Spain.*

**Dr. Mohamed Hamed Yassien**  
*National Institute of Oceanography and Fisheries, Suez  
branch,  
P.O. Box (182), Suez,  
Egypt.*

**Dr. Abhay Bhalchandra Thakur**  
*2/9 Mai Krupa Sagar Society Opp. Catering College  
Veer Savarkar Marg Dadar, Mumbai -400 028  
Maharashtra,  
India.*

**Dr. Riaz Ahmad**  
*Department of Zoology  
Aligarh Muslim University  
Aligarh- 202002, (UP)  
India.*

# International Journal of Fisheries and Aquaculture

Table of Contents: Volume 7 Number 3 March 2015

## ARTICLES

---

### Research Articles

- Food and feeding habits of *Mugil cephalus* (Linnaeus, 1758) in Elechi Creek, Niger Delta, Nigeria** 25  
Jamabo, N. A. and Maduako, N. C.
- Immunostimulatory and anti-oxidative properties of corn silk from *Zea mays* L. in Nile tilapia, *Oreochromis niloticus*** 30  
Elena S. Catap, Maria Rexie R. Jimenez and Ma. Patricia B. Tumbali

Full Length Research Paper

## Food and feeding habits of *Mugil cephalus* (Linnaeus, 1758) in Elechi Creek, Niger Delta, Nigeria

Jamabo, N. A.\* and Maduako, N. C.

Department of Fisheries, Faculty of Agriculture, University of Port-Harcourt, P. M. B. 5323, Port-Harcourt, Rivers State, Nigeria.

Received 11 November, 2014; Accepted 27 January, 2015

The present study is aimed to provide information on the abundance of natural foods needed by *Mugil cephalus* in the Elechi Creek. The natural food of *M. cephalus* in the Elechi Creek was studied from stomach contents of the fish. The stomach contents were analyzed using two methods; the frequency of occurrence and numerical methods. Plant materials, diatoms, algae and dinoflagellates constituted its main food. Plant materials were found to be the most preferable food where it occurred in 67.0% of the examined fish. Annelids, fish larva, fish parts, insect parts and crustaceans comprised the food of animal origin. Sand/mud and organic matter occurred in about 11.0 and 73.0% of the examined stomachs respectively. These results indicate that *M. cephalus* is omnivorous plant materials were the most abundant food items by numerical and occurrence methods in the gut of the fish species.

**Key words:** Algae, food items, frequency of occurrence, numerical method, stomach content.

### INTRODUCTION

The study of the food and feeding habit is useful and fundamental to understand the functional role of the fish within its ecosystem. The family Mugilidae is among the commercially important fishes occurring in Elechi Creek, Rivers State. They consist of seventeen genera and eighty species which are widely distributed along the brackish estuaries and coastal lagoons. They are popular, well accepted and form a large proportion of the diets of the rural communities in coastal areas in Nigeria. *Mugil cephalus* occurs worldwide from approximately 42° N to 42° S Latitude (Bok, 1979; Render et al., 1995), where it inhabits estuarine intertidal freshwater and coastal marine habitats. In the western Atlantic Ocean, *M. cephalus* ranges from Cape Cod to Brazil, including

the Gulf of Mexico, Caribbean, and West Indies (Fagade and Olaniyan, 1998). Adults and juveniles of grey mullets are hardy, euryhaline, eurythermal and not competitor for food among its species. School occurs in shallow coastal waters; they enter lagoons and estuaries to feed (Rheman et al., 2002). In estuarine waters, they feed on detritus, diatoms, algae and microscopic invertebrates (McDonough and Wenner, 2003). Bishop and Miglarea (1978) found that the principal food sources of adult mullet are detritus and epiphytic algae. However, they also observed *M. cephalus* feeding opportunistically on swarming polychaetes of the *Nereis* genus. The species is heterotrophic, adults feed primarily on detritus. Mulletts constitute a large proportion of the catches by artisanal

\*Corresponding author. E-mail: nenejamabo@yahoo.com

Author(s) agree that this article remain permanently open access under the terms of the [Creative Commons Attribution License 4.0 International License](https://creativecommons.org/licenses/by/4.0/)

**Table 1.** Analysis of food items stomach of *M. cephalus* by numerical and frequency of occurrence.

Food items	Numerical method		Occurrence method	
	No	%	No	%
Algae	179	19.40	45	15.68
Diatoms	229	24.81	23	8.01
Plant materials	281	30.44	67	23.35
Crustaceans	39	4.23	14	4.88
Fish parts	16	1.73	9	3.14
Insect parts	9	0.97	7	2.44
Annelids	3	0.32	2	0.70
Sand / mud	-	-	11	3.83
Organic matter	-	-	73	25.44
Dinoflagellates	163	17.65	33	11.50
Fish larva	4	0.43	3	1.05

and subsistence fishermen in lagoons and rivers and generate a high market value in demand because of their taste. This paper describes the diet and feeding habits of *M. cephalus* based on the examination of stomachs collected.

## MATERIALS AND METHODS

### Study area

The Elechi Creek also known as Elechi, Omo Ema Creek is located in Rivers State, Niger Delta Nigeria. The creek lies between longitude 6° 45'E and 7° 20'N and latitude 4° 38'N and 5° 5'E South-West of Port Harcourt. The creek is tide dominated embayment with little fresh water input and is characterized by extensive mangrove swamps, tidal flats, influenced by semi-diurnal tidal regime (NEDECO, 1961).

The vegetation consists of mangrove forest dominated by the red mangrove *Rhizophora racemosa* and *Rhizophora mangle*. In some areas, the white mangrove *Avicennia africana* is interspersed with *Nypa fructicans*. Its vegetation provides logs of wood for domestic and building purposes. The low inter-tidal zone is usually bare of vegetation, with clay, peat and sand deposit. The area is surrounded by numerous water fronts' residential houses. The surrounding terrestrial environment is marked by various human activities such as saw milling of timber.

### Sampling procedure

The food and feeding habits of *M. cephalus* was studied by examining the stomach contents. Fish samples were collected with gill and cast nets. The nets used were between 30 and 60 mm mesh size. Specimens of *M. cephalus* were collected fortnightly from the artisanal fishermen. Fish specimens were transported to the University of Port Harcourt, Faculty of Agriculture laboratory to investigate the food and feeding habits.

The fish samples were preserved in 10% formalin solution to avoid deterioration. Each fish specimen collected was given a registration number which indicates date of capture, weight and length. Standard and total lengths (in centimeter) were measured using a measuring board while the weights (in grammes) were taken to the nearest gram using a sensitive electrical balance, MP 2003 model. The stomachs were removed intact by cutting above

the cardiac and below the pyloric sphincters and preserved in a vial with 4% formalin. Dissection entailed making an incision above the longitudinal axis of the stomach and intestine. The stomach contents were emptied into a Petri dish and identified to the lowest taxonomic level. Identification entailed first sorting by eye for larger food substances. Secondly the stomach contents were dropped on the slides with the aid of a dropping pipette and observed under a light microscope (with variable magnifications up to X 40) examinations.

Food was regarded as total ingested matter and was analysed using the frequency of occurrence and the numerical methods (Bagenal, 1978; Hyslop, 1980). In the frequency of occurrence each food item was used to indicate the proportion of fishes eating a particular food item. The merits and limitations of these methods have been discussed by Dunn (1954), Crips (1963) and Fagade and Olaniyan (1972).

$$\text{Frequency of occurrence} = X/Y \times 100\%$$

Where, X = number of stomach where each food item is present, Y = number of stomachs for the experiments.

In numerical methods, the number of individual in each category is recorded for all stomachs and the total is expressed as a proportion.

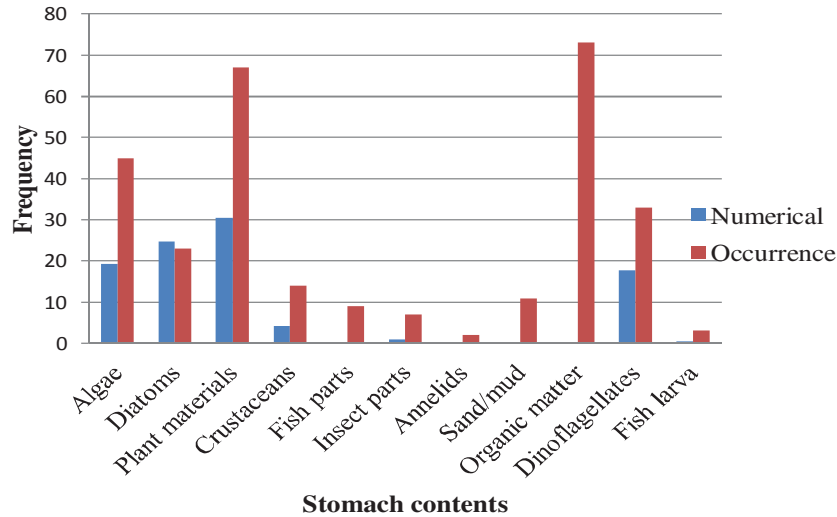
## RESULTS

### Food composition

Table 1 show the list of food items found in the stomach of *M. cephalus*, while the summary of the items are illustrated in Figure 1. The relative contributions of the food items are expressed by frequency of occurrence and numerical methods. A total of 100 stomachs were randomly examined. Eleven major items constituted the diet of *M. cephalus*, four forming the dominant stomach content.

### Food composition for juvenile's *M. cephalus*

In the numerical analysis plant materials were dominant



**Figure 1.** Summary of the stomach contents of *M. cephalus* from the Elechi Creek.

and composed of 30.44% of the items in the stomach, diatoms made up 24.81%, while algae made up 19.40%, dinoflagellates 17.65%, crustaceans 4.23%, fish parts and insect parts 1.73 and 0.97%. Fish larva and annelids were the least with 0.43 and 0.32% respectively.

Food items were contained in one hundred stomachs and organic matter (detritus) and sand particles were present in 25.44%, plant materials were contained in 23.35% of the stomach, algae 15.68%, dinoflagellates 11.50%, diatoms 8.01%, crustaceans 4.88%, fish and insect parts had 3.14 and 2.44%. The least food items of 1.05 and 0.70% were found in fish larva and annelids respectively.

The result of the stomach content of the various size groups is presented in Table 2. Plant materials constituted the most consumed food items fed on in the three size groups both by numerical method (30.91, 33.84 and 25.0) and by frequency of occurrence (33.33, 33.84 and 17.89) methods. Detritus (organic matter) constituted the most occurring item in the stomachs of the three size groups of *M. cephalus* from the Elechi Creek by occurrence (37.17, 24.32, and 17.89). Plant materials were the next in occurrence to the detritus in number of stomach where they are found in the three size groups. The medium size groups consumed more food items than the other size groups.

Variation in empty stomach by size group (Table 3) indicated that the small size group of *M. cephalus* had the highest number of empty stomachs (8.51%), while the medium sized group had (6.25%) and the large sized group had no empty stomach.

## DISCUSSION

The food composition of the species in Elechi Creek

shows a high food richness as the specimens showed high trophic flexibility by the wide variety of food items consumed. *M. cephalus* is diurnal feeder, consuming mainly zooplanktons, dead plant matter and detritus. In this study *M. cephalus* foods varied from microscopic items such as diatoms to macroscopic ones such as annelids and fish parts. This is an indication that it has advantages of using large aquatic resource to its advantage. This high trophic flexibility ensures a constant energy source which is necessary for its large population (King, 1988). Its occurrence maybe due to this reason; the fish was able to shift from one form of food to another depending on the seasons and availability of the foods. Availability of food organisms are often cyclic and maybe due to their life histories, climate, or environmental conditions. During this study, empty stomach was recorded between the months of August (12.5%) and September (5.0%).

Presence of plants presumed the species as an herbivore, while the availability of invertebrates showed it as a carnivore, fish parts as a part of its diet was an indication of its piscivorous ability. Presence of organic matter and sand/mud in reasonable number mean the species was both detritus and benthic feeder food is ingested from bottom substrate.

In relation to size, the trend among the three size groups of the stripped mullets from the Elechi Creek changes. The juveniles fed mainly on plant materials, algae, dinoflagellates, diatoms and crustaceans, while the adults predominantly fed on plant materials, algae, dinoflagellates, diatoms, crustaceans, annelids, fish parts, insect parts, sand/mud and organic matter. The high dietary importance of sand/mud and detritus may be due to the marginal mangrove vegetation growing on water-logged deposits of soft mud and clay-silt sediment.

The decomposing leaves and other biogenic materials



**Table 2.** Stomach contents of *M. cephalus* by size groups from Elechi Creek.

Stomach contents	Small-sized (11.2-13.cm)			Medium-sized (13.7-18.5cm)			Large-sized (18.6-24.7cm)					
	Numerical method		Occurrence method	Numerical method		Occurrence method	Numerical method		Occurrence method			
	No	%	No	%	No	%	No	%	No	%		
Algae	42	24.6	11	14.1	83	18.4	21	18.9	54	18.0	13	13.7
Diatoms	26	15.2	3	3.9	118	26.1	11	9.9	85	28.3	9	9.5
Plant materials	53	30.9	26	33.3	153	33.8	24	21.6	75	25.0	17	17.9
Crustaceans	9	5.3	2	2.6	18	4.0	7	6.3	12	4.0	5	5.3
Fish parts	-	-	-	-	5	1.1	3	2.7	11	3.7	6	6.3
Insect parts	-	-	-	-	2	0.4	1	0.9	7	2.3	6	6.3
Annelids	-	-	-	-	1	0.2	1	0.9	2	0.7	1	1.1
Unidentified items	-	-	-	-	-	-	3	2.7	-	-	8	8.4
Organic matter	-	-	29	37.2	-	-	27	24.3	-	-	17	17.9
Dinoflagellates	41	24.0	7	9.0	71	15.7	15	13.5	51	17	11	11.6
Fish larva	-	-	-	-	1	0.2	1	0.9	3	1	2	2.1

**Table 3.** Variation in empty stomach by size of *M. cephalus* from Elechi Creek.

Size/standard length (cm)	Number examined	Number with empty stomach	% Empty stomach
Small-sized (11.2 - 13.6)	47	4	8.51
Medium-sized (13.7 - 18.5)	32	2	6.25
Large-sized fish (18.6 - 24.7)	21	-	-

washed into the swamp ensure a constantly enriched nutrient pool for algae often seen growing extensively on the exposed mud surfaces.

The adults had more of sand/mud and organic matter in their stomach because they become benthic feeder as they grow. This change in diet with growth not only offers a wider range of food resources to the species but also reduces possible competition between the adults and juveniles to some extent. Seasonally variation in the feeding habit was not recorded as the study was carried out only in the rainy season. The

success of mullets according to Kurian (1975) and Payne (1976) also lies in their feeding habits and the abundance of their food. Presence of detritus and sand/mud in large quantities in the stomach of the fish does not presume that these items were nutritional better than other food items that appeared in lesser number. Sand/mud has no nutritional property but they might have been picked up along with other food items. Its presence in the gizzard like stomach may assist the fish in digestion process.

In comparison with other studies, this result confirmed to the study on food and feeding habits

with reviews from Akpan and Ubak (2005) and Lawson and Jimoh (2010), on the diets of mullets. However, from the study, major food of *M. cephalus* in the Elechi Creek where diatoms, algae, plant materials, dinoflagellates, sand/mud, organic matter, crustaceans, insect parts, fish parts and annelids.

#### Conflict of Interest

The authors have not declared any conflict of interest.

## REFERENCES

- Akpan AW, Ubak RG (2005). Aspects of the trophic ecology of *Mugilidae* (Valenciennes, 1830) in the Cross River Estuary, Nigeria. *J. Aquat. Sci.* 20(1):53-62.
- Bagenal TB (1978). Methods for assessment of fish production in fresh water, Blackwell Scientific publications, Oxford.
- Bok AH (1979). The distribution and ecology of two mullet species in some freshwater rivers in the eastern cape, South Africa. *J. Limnol.* 5:97-102.
- Bishop JM, Miglajese JV (1978). Carnivorous feeding in adult striped mullet. *Copeia*, pp. 705-709.
- Crips DT (1963). A preliminary survey of brown trout (*Salmo trutta* L.) and bullheads (*Cottasgobia* L.) in high altitude becks. *Salmon Trout Mag.* 167:45-59.
- Dunn DR (1954). The feeding habits of some of the fishes and some members of the bottom fauna of Llyn Tegid (Bala Lake), Merionethshire. *J. Anim. Ecol.* 23:224-233.
- Fagade SO Olaniyan CIO (1972). The Biology of the West African shad *Ethmalosa fimbriata* (Bowditch) in the Lagos Lagoon, Nigeria. *J. Fish Biol.* 4:519-533.
- Fagade SO, Olaniyan CIO (1998). The food and feeding interrelationship of the fishes in Kainji Lake. *J. Fish Biol.* 5:205-227.
- Hyslop EJ (1980). Stomach content analysis: A review of methods and their Application. *J. Fish Biol.* 17:411-429.
- King RP (1988). Observations on *Liza falcipinus* (Valenciennes, 1836) in Bonny River, Nigeria. *Hydrobiol. Tropics* 21(3):63-70.
- Kurian A (1975). The identification of Grey mullet species by disc Electrophoresis. *Aquaculture* 5:99-106.
- Lawson EO, Jimoh AA (2010). Aspects of the biology of grey mullet, *Mugil cephalus*, in Lagos Lagoon, Nigeria. *AACL Bioflux.* 3(3):181-193.
- McDonough CJ Wenner CA (2003). Growth, recruitment and abundance of juvenile *Mugil cephalus* in South Carolina estuaries" *Fish. Bull.* 101:343-357.
- NEDECO (1961). The waters of the Niger Delta: Reports of an investigation by NEDECO (Netherlands engineering consultants)" The Hague.
- Payne AI (1976). The relative abundance and feeding of the grey mullet species occurring in an estuary in Sierra Leone, West Africa. *Marine Biol.* 35:227-286.
- Render JH Thompson BA Allen RL (1995). Reproductive development of stripped mullet in Louisiana estuarine waters with notes on the applicability of reproductive assessment methods for isochronal species. *Trans. Am. Fish. Soc.* 124:26-36.
- Rhemam S, Islam ML Shah MMR Mondal S Alan MJ (2002). Observation on the fecundity and Gonadosomatic Index (GSI) of Grey mullet, *Liza parsia* (Ham.). *Online J. Biol. Sci.* 2(10):690-693.

Full Length Research Paper

# Immunostimulatory and anti-oxidative properties of corn silk from *Zea mays* L. in Nile tilapia, *Oreochromis niloticus*

Elena S. Catap<sup>1,2\*</sup>, Maria Rexie R. Jimenez<sup>1,2</sup> and Ma. Patricia B. Tumbali<sup>1</sup>

<sup>1</sup>Institute of Biology, College of Science, University of the Philippines, Diliman, Quezon City, Philippines 1101, Philippines.

<sup>2</sup>Natural Sciences Research Institute, University of the Philippines, Diliman, Quezon City, Philippines 1101, Philippines.

Received 29 September, 2014; Accepted 2 February, 2015

Corn silks are threads found in the maize plant (*Zea mays*) and traditionally used to treat urological infections and disorders. Corn silk is also known to possess nutrients and volatile compounds. However, this material is often disregarded and unused. This study therefore investigated the potential use of corn silk in aquaculture through its protective capacity in matured Nile tilapia (*Oreochromis niloticus* (L.)) by measuring some nonspecific immune parameters (phagocytosis, production of reactive oxygen species, and plasma lysozyme level) in experimental *Aeromonas hydrophila*-challenged fish. The anti-oxidative property of corn silk was also investigated using paracetamol-induced hepatic toxicity in order to measure oxidative stress (malondialdehyde or MDA). Based on the results, phagocytosis was significantly higher in *A. hydrophila* infected fish fed with corn silk-coated feeds than in fish from the negative (PBS-injected) and positive control (*Aeromonas hydrophila* infected) treatments. Lysozyme level was also higher in corn silk-fed fish, but it was not significantly different from the positive control fish (*A. hydrophila* infected fish). Reactive oxygen species (ROS) was higher in corn silk-fed fish than the positive control fish but it was not statistically significant. MDA levels were significantly higher in paracetamol-treated fish than paracetamol-corn silk treated group. The results showed the potential immunostimulatory and antioxidant role of corn silk in Nile tilapia, but further studies are required to fully understand its mechanism of action and its full use in aquaculture.

**Key words:** Corn silk, lipid peroxidation, *Aeromonas hydrophila*, *Oreochromis niloticus*, immunostimulation.

## INTRODUCTION

Presently, aquaculture is a fast growing food production industry that contributes nearly 50% of the annual fisheries production (FAO, 2012). In the Philippines, tilapia is the second most cultured fish species after milkfish. However, annual production of Nile tilapia is usually

affected by episodes of high mortality mostly due to bacterial infections that could be attributed to very intensive culture practices and sometimes aggravated by seasonal effects of low environmental temperatures. One of the opportunistic bacteria that infect cultured Nile tilapia

\*Corresponding author. E-mail: [elenacatap@yahoo.com](mailto:elenacatap@yahoo.com)

Author(s) agree that this article remain permanently open access under the terms of the [Creative Commons Attribution License 4.0 International License](https://creativecommons.org/licenses/by/4.0/)

is *Aeromonas hydrophila* which causes septicemic infections in the fish (Cipriano, 2001). These pathogens are ubiquitous and are thus found in a variety of aquatic environments. To control these bacterial infections, antibiotics are mainly used.

However, antibiotic use may lead to the emergence or development of antibiotic-resistant bacteria, thus safer and more effective alternatives should be used (Pachanawan et al., 2008). Studies on the use of immunostimulants derived from natural bioactive products are presently gaining importance as an option instead of using chemotherapeutants and antibiotics. Immunostimulants enhance the non-specific immune responses (innate immunity) as well as the specific immune response mechanisms (adaptive or acquired immunity) of a certain organism (Anderson, 1992). Other immunostimulants like chitin, chitosan, and levamisole have been reported to enhance the non-immune responses of common carp (*Cyprinus carpio*) which led to a higher percentage survival of the fish (Gopalakannan and Arul, 2006).

Corn silk (*Zea mays*) is traditionally used in the treatment of cystitis, edema, gout, kidney stones, nephritis, and urological disorders (Ebrahimzadeh et al., 2008). It is also used as an anti-diabetic agent since it reportedly counteracts hyperglycaemia (Guo et al., 2009). It has phenolic compounds such as anthocyanins, vanillic acid, and protocatechuic acid which are reportedly responsible for its antioxidant capacity (Ebrahimzadeh et al., 2008). Corn silk extract has also shown great potential when it comes to prevention of diseases involving overproduction of radicals (Liu et al., 2011). Administration of natural products may be done via bathing, injection, or oral administration, the latter of which is the least stressful of all the three routes (Harikrishnan et al., 2011).

This study was therefore undertaken to investigate the protective role of dietary corn silk supplements against *A. hydrophila* infection and induced hepatic damage by measuring some nonspecific immune parameters and lipid peroxidation activity as indicator of oxidative stress.

## MATERIALS AND METHODS

### Fish and experimental design

Mixed sex *Oreochromis niloticus* obtained from the Southeast Asia Fisheries Development Center, Freshwater Fisheries Station, Binangonan, Rizal and Philippines were used. Prior to the experiments, the fish were acclimatized in rectangular tanks with recirculating water system. They were fed daily at a feeding rate of 3% of the total body weight with commercial grow-out tilapia pellets. Regular water change was done during the pre-experimental period. For the experiments, three groups of tilapia, 20 fish each ( $58.9 \pm 1.76$  g, 180 fish total), were designated: a) negative control, which consisted of healthy tilapia fed with commercial pelleted feeds; b) positive control, which consisted of tilapia infected with *A. hydrophila* and fed with commercial tilapia feed pellets; and c) corn silk treated group, which consisted of tilapia infected with *A.*

*hydrophila* and fed with the corn silk-coated tilapia feed pellets.

In order to determine the antioxidant property of corn silk, another experimental set-up consisted of four groups of tilapia ( $42.5 \pm 0.98$  g; 80 fish) that were designated as: a) negative control, which consisted of tilapia fed with commercial pelleted tilapia feeds; b) positive control, which consisted of tilapia fed with feeds coated with paracetamol; c) paracetamol and silymarin treated group, which consisted of tilapia fed with a mixture of paracetamol and silymarin-coated fish food; and d) paracetamol and corn silk treated group, which consisted of tilapia fed with a mixture of paracetamol and corn silk extract-coated fish food.

### Fish food preparation

The powdered corn silk (1.750 g) produced by Britmix Wellness, Inc. was coated onto 500 g of commercial fish feed pellets. The powdered corn silk was mixed into a 22% gelatin solution and this mixture was coated onto the fish feed pellets. The coated feeds were air dried for 48 h and both the coated and uncoated feed pellets were stored in the refrigerator (4°C) to prevent bacterial and fungal contamination. The corn silk-coated fish food pellets were used for the fish in the first experiment. For the second experiment, four types of fish diets were used: a) for the negative control group, unsupplemented commercial fish food, b) for the positive control group, powdered paracetamol (1000 mg) was coated using gelatin onto 500 g of fish feed pellets, c) for the paracetamol-silymarin reference group, 1000 mg of paracetamol and 450 mg of silymarin (Steinbach Products, Inc.; Mandaluyong, Philippines) were coated onto 500 g fish feed pellets, and d) for the paracetamol-corn silk group, 1000 mg of paracetamol and 1000 mg of corn silk were coated onto 500 g fish feed pellets. The fish were fed at 3% body weight daily during the experimental period.

### Preparation and Injection of bacteria

*A. hydrophila* culture was obtained from the National Institute of Molecular Biology and Biotechnology (BIOTECH) of the University of the Philippines - Los Baños. The bacterial inoculant was prepared through serial dilution and spread plate technique. Final bacterial concentration was adjusted to  $6.2 \times 10^5$  CFU ml<sup>-1</sup>. A 0.1 ml of the bacterial suspension was injected intramuscularly near the lateral line of the pectoral region of the fish from the positive control group and corn silk treated group of the first experimental set-up. Phosphate buffer saline (PBS) pH 7.2 with the same volume was injected into the negative control tilapia group. Bacterial infection was done after a 30 day feeding period.

### Sample collection

For the immune response experiments, the fish in each treatment tanks were sacrificed at day 7 post-infection. Each fish was immobilized with a blow in the head. Blood was extracted from the midventral caudal peduncle. Plasma was obtained from the extracted blood by centrifuging at 10,000 rpm for 5 min at 4°C. Head kidney were dissected out and immersed in Petri dishes with cold supplemented fish physiological saline (FPS). For the antioxidant experiments, fish were sacrificed from each of the four tanks at day 16 of the experimental period. The livers were removed and immersed in PBS to be used in lipid peroxidation assay.

### Lysozyme assay

Dilutions of hen egg white lysozyme (HEWL) was used as standard

and prepared in phosphate citrate buffer (pH 5.8). *Micrococcus lysodeikticus* solution (75% w/v) buffered to pH 5.6 in phosphate citrate was mixed to HEWL standard dilution or tilapia plasma in a microtiter plate 7:1 ratio. Absorbance at 450 nm was read after 15 min using the ELISA plate reader. Using the standard curve of Vmax rates, plasma lysozyme concentrations were determined. Protein concentrations of the plasma samples were also determined.

#### Macrophage phagocytic activity assay

The head kidney samples were homogenized using a screen mesh and suspended in 3 ml supplemented Leibovitz-15 medium (L-15). The cells were centrifuged at 400 rpm for 5 min in room temperature. The pelleted cells were washed twice with supplement L-15. After washing, the cells were suspended in FPS and the cell count and viability was determined by staining the cells with trypan blue (1:9), utilizing the trypan blue exclusion method. The cell concentration was adjusted to  $10^6$  viable cells  $\text{ml}^{-1}$ . Opsonized yeast cells were used as feeds to phagocytes, approximately with  $10^6$  yeasts  $\text{ml}^{-1}$  in PBS, prepared and mixed with one (1.0) ml of 0.8% Congo red dye. The yeast suspension was then autoclaved and washed with an equal volume of PBS until the excess Congo red dye was removed.

The suspension was centrifuged at 400 rpm for 5 min. The cells obtained were suspended again in PBS. The prepared yeast cells were added to the cell suspension from each sample at 2:1 ratio. An aliquot sample of 20  $\mu\text{l}$  was smeared onto a glass slide after one hr of incubation at room temperature. The smears were air dried and fixed with 95% ethanol. After 24 h, the films were stained with 1% eosin for one min, rinsed with distilled water, and dipped in Giemsa stain for two minutes. These were air dried for 24 h. Cover slips were mounted onto the glass slides using Entellan. The smears were observed under the oil immersion objective. Percentage of active phagocytes was recorded from a hundred phagocytes that were counted in representative areas of the slide. Data were expressed as mean percentages of active phagocytes.

#### Reactive oxygen species (ROS) production assay

The assay for the assessment of superoxide anion produced outside the mitochondria was done according to the protocol of Zelickoff (1996). Head kidney macrophages were obtained from the previous preparation and adjusted to a cell concentration of  $4 \times 10^6$  cells  $\text{ml}^{-1}$ . Four microcentrifuge tubes were labeled (1 to 4), and cells ( $125 \mu\text{l}$  of  $10^6$  cells  $\text{ml}^{-1}$ ) were added to each of these four tubes containing 250  $\mu\text{l}$  ferricytochrome solution (final concentration =  $2 \text{ mg ml}^{-1}$ ) and 62.5  $\mu\text{l}$  of bovine superoxide dismutase was added to the second and fourth tubes.

Ten microliters of phorbol 12-myristate 13-acetate was added to the third and fourth tubes at a final concentration of  $2 \mu\text{g ml}^{-1}$ . Fish physiological saline solution (FPS) was added to each tube to bring up the total volume to 0.5 ml. An additional tube containing all the reagents but without the cells served as the blank. Each tube was vortexed for approximately 30 s and 200  $\mu\text{l}$  aliquot placed into the wells of a 96-well microtiter plate. The absorbance was measured at 492nm for up to an hour. Time points used for measurement include: 0, 15, 30, 45, and 60 min. The plates were incubated at room temperature, in a humid environment, between readings. The rate of superoxide anion radical production was determined from measurements taken over time, while OD readings at a single time point were used to make comparisons between different exposure groups. Change in absorbance was calculated by subtracting the mean of the "blank" wells and the wells containing SOD from the absorbance measured in the non-SOD-containing wells. By multiplying the change in absorbance by 15.87 the nmol

concentration of SOD-inhibitable superoxide anion radical was computed. Data are expressed as nmol  $\text{O}_2/2 \times 10^5$  cells/unit time.

#### Lipid peroxidation assay

The thiobarbituric acid reactive substance assay (TBARS assay) was used to measure lipid peroxidation in hepatic tissues obtained from tilapia. It measure malondialdehyde (MDA), one of the compounds formed by lipids after oxidative processes. Briefly, using a ground glass homogenizer, one gram of liver tissue was homogenized in 2 ml of PBS. An aliquot of 0.5 mL of the homogenate was transferred in clean test tubes and 2.5 ml of trichloroacetic acid (TCA) and 1 ml of thiobarbituric acid (TBA) was added to all test tubes and then mixed through vortexing.

The samples were then subjected to a hot water bath for 30 min. The samples were allowed to cool and then 4 ml of butanol was added to each of the test tubes. The samples were vortexed and the organic layer was removed and placed in centrifuge tubes. The organic layer was centrifuged at 3000 rpm for 10 min at room temperature. The absorbance was read at 532 nm using tetramethoxypropane was used as the standard. Data were expressed in  $\mu\text{mol MDA. mg protein}^{-1}$ .

#### Protein content determination

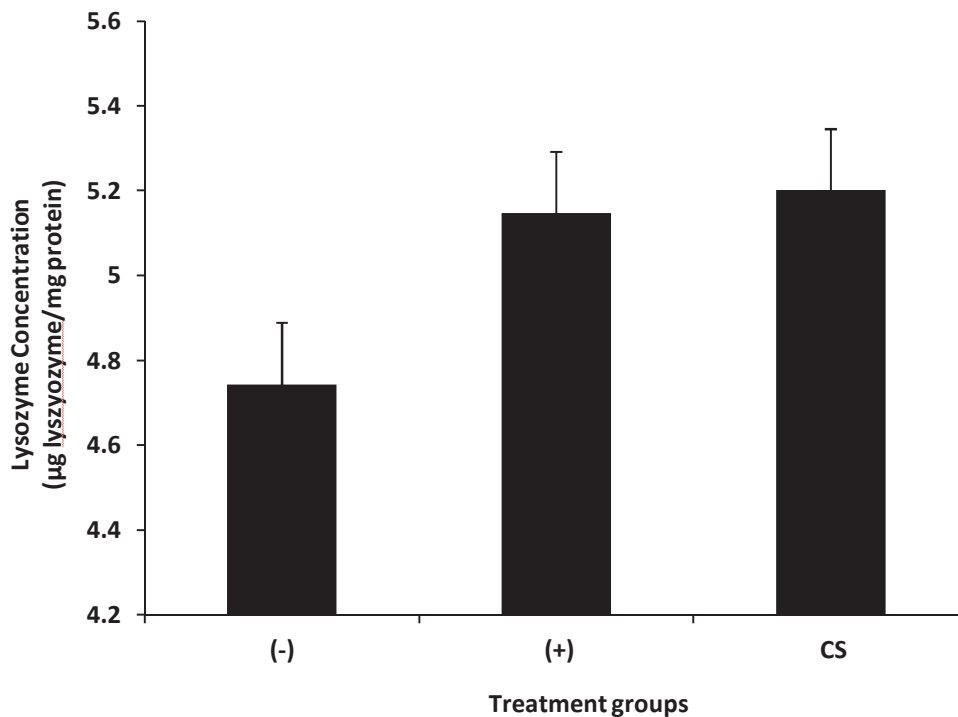
Protein content was determined with the use of the BIO-Rad protein assay kit. The dye was prepared by diluting one part of the concentrated dye with four parts of deionized water. The filtrate was then collected. Dilutions of BSA were used as standard. 10  $\mu\text{l}$  of each standard and sample solution were added to 96 well plates. 200  $\mu\text{l}$  of the diluted dye was then added to all wells. The plate was incubated for 5 min at room temperature and absorbance was read at 595 nm with the use of an ELISA plate reader. Data were expressed as  $\text{mg ml}^{-1}$ .

#### Data analysis

All of the data were analysed for normality of distribution using Shapiro-Wilk test and homogeneity of variance prior to one way ANOVA test at  $P < 0.05$ . Kruskal-Wallis test was used as non-parametric test for data with non-normal distribution. Comparison of the data was employed by using Least Significance Difference test for homogenous data sets and Games-Howell test for non-homogeneous data. MDA concentration values were analysed using Mann-Whitney U test.

## RESULTS AND DISCUSSION

The efficiency of an immunostimulant is evaluated by testing its ability to protect the fish against pathogens, and also by measuring the immune response produced (Galindo-Villegas and Hosokawa, 2004). In the present study, the protective effects of dietary corn silk supplementation were determined in Nile tilapia. In order to determine the immune enhancing property of corn silk, fish were fed for 30 days with corn-silk coated feed pellets and then inoculated with *A. hydrophila*. Subsequently, phagocytic activity, plasma lysozyme levels and production of reactive oxygen species were measured as indicators of nonspecific immune response. Experiments were also undertaken in order wherein



**Figure 1.** Plasma lysozyme levels from Nile tilapia (*Oreochromis niloticus*). Data are presented as mean  $\pm$  SEM. (-), negative control, (+), positive control, CS, corn silk-treated group.

hepatic injury was induced through paracetamol-corn silk treatment to determine if corn silk could ameliorate oxidative stress.

Corn silk is composed of stigmas and styles of the *Zea mays* (maize) plant, and it has been used in traditional Chinese medicine for the treatment of various ailments (Ren et al., 2009). It has been reported that corn silk is an excellent source of many bioactive compounds such as flavonoids, saponin, alkaloids, tannins, phytosterols, allantoin, vitamin E and K, etc. (Hu and Deng, 2011; Ren et al., 2013). It is possibly due to these bioactive components that research studies in corn silk were undertaken to identify its relevance to human health, which includes its reported immune enhancing effects. Similarly, fish health researchers have continually searched for bioactive compounds that could be used to enhance fish immunity and protection against various pathogens. Best known immune stimulants are glucans and lipopolysaccharides, and synthetic compounds, animal and plant extracts and vitamins that usually target the nonspecific immunity of cultured fish species (Ardo et al., 2008).

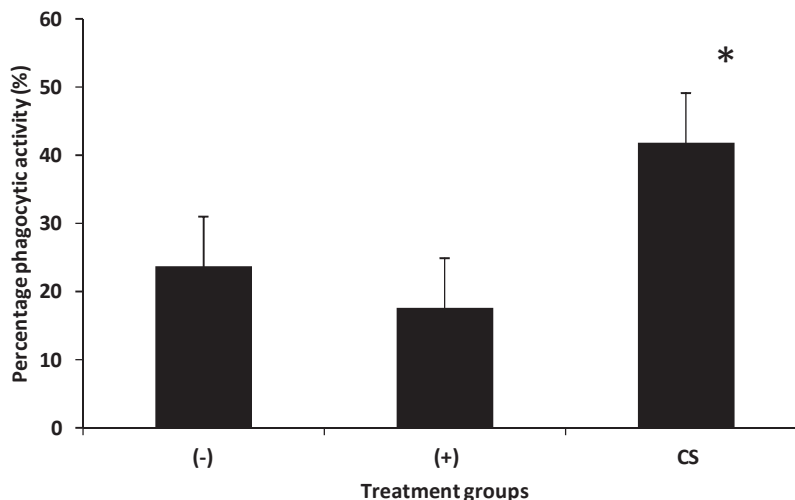
Based on the results, higher lysozyme level was exhibited by the group of fish which were fed with corn silk coated feed pellets and infected with *A. hydrophila* (Figure 1) but the increase was not significantly different from that of the positive control group. It is likely that the concentration of the powdered corn silk added may not have been potent enough to raise the lysozyme levels in

tilapia significantly. Alternately, the duration of the feeding period prior to the bacterial challenge could have been longer in order to boost the immune system.

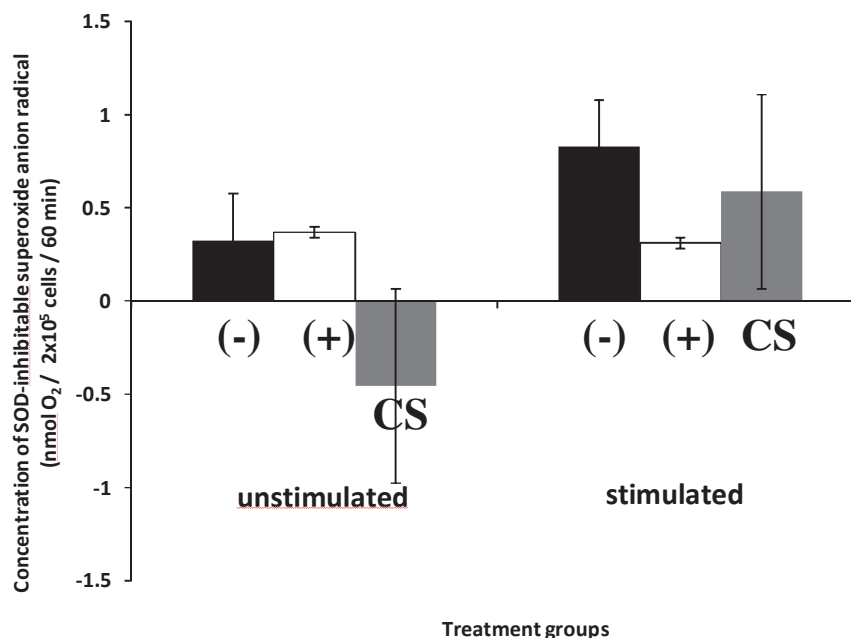
However, it is likely that this increase in lysozyme level could also aid in the destruction of *A. hydrophila*. It is also worthy to note that bioactive compounds from corn silk could also inhibit bacterial infection. In a study by Nessa et al. (2012), antimicrobial activities of corn silk extracts and bioactive compounds were compared with that of gentamycin, and they found out that the extracts and flavonoids were significantly more sensitive against a number of bacteria.

Phagocytosis is the most primitive immune response mechanism, and basically involves the ingestion of a pathogen by macrophages or neutrophils. Phagocytic activity has been significantly enhanced by the corn silk treatment in this study. The corn silk treated group has statistically significant phagocytic activity than the positive and negative control groups (Figure 2). These results likewise indicate a potential immunostimulatory effect of corn silk in *A. hydrophila* infected tilapia. This response has also been exhibited by some fish species when treated with bioactive extracts. Yin et al. (2009) reported increase in phagocytosis in carp when treated with *Ganoderma lucidum* and *Astragalus radix*. They attributed these mainly to the polysaccharides, monosaccharides, flavonoid and alkaloid contents of both herbs.

Production of ROS is considered as an important



**Figure 2.** Percentage phagocytic activity of head kidney phagocytes. Data are presented as mean  $\pm$  SEM. \*- CS, corn silk treated group significantly different ( $P < 0.05$ ) from the positive control.

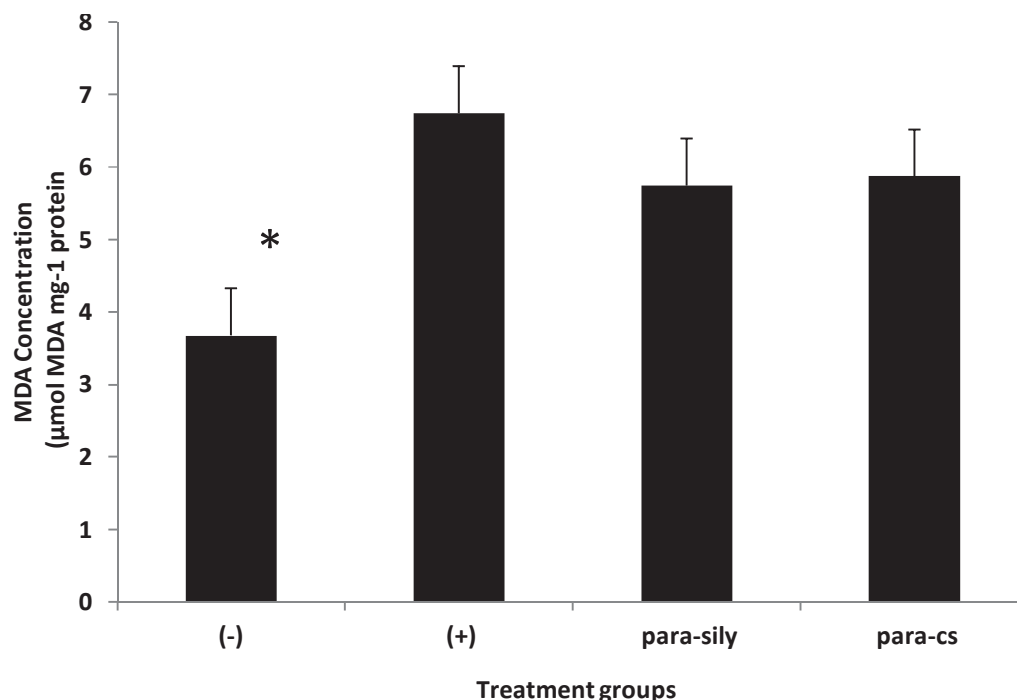


**Figure 3.** Concentration of SOD-inhibitable superoxide anion radical (nmol O<sub>2</sub><sup>-</sup>/ 2x10<sup>5</sup> cells/60 min) of head kidney macrophages in Nile tilapia. Data are presented as mean  $\pm$  SEM. (-) negative control, (+) positive control, CS- corn silk-treated group.

microbial killing mechanism (intracellular and extracellular) in vertebrates. Animals have inherent enzymes (antioxidants) to detoxify these anions to counteract the possible adverse effects in normal cells and tissues. Antioxidants include superoxide dismutase, catalase, glutathione S-transferase, glutathione peroxidase, vitamin E components such as  $\alpha$ -tocopherol and  $\gamma$ -tocopherol. One study suggested that  $\alpha$ -terpineol,

citronellol, and eugenol, are some of the main compounds involved in the antioxidant action of corn silk (El-Ghorab et al., 2007). More recent studies relate flavonone glycosides as potent antioxidants (Hu and Deng, 2011; Ren et al., 2013).

This study confirmed the ROS scavenging capacity of corn silk especially in unstimulated head kidney cells (Figure 3) where ROS production was greatly inhibited.



**Figure 4.** MDA concentrations in Nile tilapia at day 16 of paracetamol treatment. Data are presented as mean values  $\pm$  SEM. \* (-) group significantly different from the (+) and CS groups ( $P < 0.05$ ). (-), negative control fish, fed with unsupplemented diet; (+), positive control fish, diet with paracetamol; para-sily, paracetamol and silymarin-treated group, diet with paracetamol and silymarin; para-cs, paracetamol and corn silk-treated group, diet with paracetamol and corn silk.

This effect was also shown by lower ROS levels obtained in PMA-stimulated cells from corn silk-treated fish. The scavenging activity and antioxidative capacity (anti-lipid peroxidation) have been recently confirmed also by Hu and Deng (2012) and Ren et al. (2013). It should be taken note of that ROS are actually produced by macrophages as microbial killing mechanisms. Kim et al. (2005) reported that corn silk activates murine macrophages to produce cytokines and enzymes that are important in regulating normal physiological functions inflammatory response in macrophages. The data from this study likewise showed the immunostimulatory role of corn silk in macrophages from tilapia head kidney. As for oxidative stress results, the negative control fish had significantly lower MDA levels than the positive control fish as shown in Figure 4. The paracetamol-corn silk treated group has a lower concentration of MDA compared to the paracetamol-silymarin treated group, but this was not statistically different. The results showed that corn silk had comparable antioxidant effects as silymarin, and these treatments were able to alleviate, although not significantly, the oxidative effects of paracetamol on the hepatic cells. Paracetamol hepatotoxicity is highly dependent on the cytochrome P450 enzyme system. As paracetamol is metabolised at the cytochrome P450 system, it is converted to the toxic metabolite N-acetyl-P-benzenequinoneimine (NAPQI). NAPQI is a toxic free

radical that binds to proteins and fatty acids and reacts with glutathione, a natural antioxidant, eventually depleting it. Most importantly, paracetamol leads to the formation of reactive oxygen and nitrogen species that induce oxidation of cellular membranes.

In the present study, silymarin was used as reference substance since it is known to have four different flavonoids that make it a good antioxidant. Its main flavonoid component is silbin with isosilibinin, silydianin, and silychristin. It has been proven to protect against carbon tetrachloride toxicity, acetaminophen (paracetamol), phalloidin, galactosamine, and thioacetamide (Pradhan and Girish, 2006; Pradeep et al., 2007). Similarly, corn silk contains many isolates of flavonoids like myricetin, fisetin, quercetin, naringin and luteolin which have been shown to possess antioxidant and prooxidant properties of varying degrees.

However, some studies showed different flavonoid synergists as most effective in hepatoprotection. Moreover, it has been reported that corn silk effectively increases antioxidant enzyme levels such as sodium dismutase and glutathione peroxidase (Hu and Deng, 2011; Nurhanan et al., 2012). In a study by Liu et al. (2011), two flavones glycosides were isolated from the n-butanol extracts of corn silk that exhibited very high antioxidant and free-radical scavenging activities. The present study confirmed the potential of corn silk as a



potent antioxidant in a cultured fish species. This could be relevant in feed formulation where high concentrations of lipids or fatty acids are used, and thus necessitates the addition of antioxidants. However, the appropriate dietary dosage in fish still need to be further investigated. Likewise, further immune-based trials are needed to maximize fully the protective property of corn silk against fish pathogens.

### Conflict of Interest

The authors have not declared any conflict of interest.

### ACKNOWLEDGEMENTS

The authors are grateful to the University of the Philippines Diliman - Natural Sciences Research Institute (NSRI) which has provided financial support for the study. They are also grateful to the National Institute of Molecular Biology and Biotechnology (BIOTECH) of the University of the Philippines - Los Baños for the *Aeromonas hydrophila* culture used in the research project.

### REFERENCES

- Anderson DP (1992). Immunostimulants, adjuvants, and vaccine carriers in fish: applications to aquaculture. *Annu. Rev. Fish Dis.* 1:281-307.
- Ardo L, Yin G, Xu P, Varadi L, Szigeti G, Jeney Z, Jeney G (2008). Chinese herbs (*Astragalus membranaceus* and *Lonicera japonica*) and boron enhance the non-specific immune response of Nile tilapia (*Oreochromis niloticus*) and resistance against *Aeromonas hydrophila*. *Aquaculture* 275:26-33.
- Cipriano R (2001). *Aeromonas hydrophila* and motile *Aeromonas* septicemias of fish. Revision of *Fish Disease Leaflet* 68 (1984) by R. Cipriano, G.L. Bullock and S.W. Pyle.
- Ebrahimzadeh M, Pourmorad F, Hafezi S (2008). Antioxidant activities of Iranian corn silk. *Turk. J. Biol.* 32:43-49.
- El-Ghorab A, El-Massry K, Shibamoto T (2007). Chemical composition of the volatile extract and antioxidant activities of the volatile and nonvolatile extracts of Egyptian corn silk (*Zea mays* L.). *J. Agric. Food Chem.* 55:9124-9127.
- Food and Agriculture Organization of the United Nations (FAO) (2012). Main Cultured Species. <http://www.fao.org/fishery/topic/13531/en>.
- Galindo-Villegas J, Hosokawa H (2004). Immunostimulants: temporary prevention of diseases in marine fish. *Avances in Nutricion Acuicola VII. Memorias del VII Simposium Internacional de Nutricion Acuicola*. <http://www.uanl.mx/secciones/publicaciones/nutricion/VII/archives/16JorgeGalindo.pdf>.
- Gopalakannan A, Arul V (2006). Immunomodulatory effects of dietary intake of chitin, chitosan and levamisole on the immune system of *Cyprinus carpio* and control of *Aeromonas hydrophila* infection in ponds. *Aquaculture* 255:179-187.
- Kim KA, Shin H-H, Choi SK, Choi H-S (2005). Corn silk induced cyclooxygenase-2 in murine macrophages. *Biosci. Biotechnol. Biochem.* 69:1848-1853.
- Guo J, Liu T, Han L, Liu Y (2009). The effects of corn silk on glycaemic metabolism. *Nutr. Metabolism* 6:47.
- Harikrishnan R, Balasundaram C, Heo MS (2011). Impact of plant products on innate and adaptive immune system of culture finfish and shellfish. *Aquaculture* 317:1-15.
- Hu Q, Deng Z (2011). Protective effects of flavonoids from corn silk on oxidative stress induced by exhaustive exercise in mice. *Afr. J. Biotech.* 10:3163-3167.
- Liu J, Wang C, Wang Z, Zhang C, Lu S, Liu J (2011). The antioxidant and free-radical scavenging activities of extract and fractions from corn silk (*Zea mays* L.) and related flavone glycosides. *Food Chem.* 126:261-269.
- Nessa F, Ismail Z, Mohamed N (2012). Antimicrobial activities of extracts and flavonoid glycosides of corn silk (*Zea mays* L.). *Int. J. Biotechnol. Wellness Ind.* 1:115-121.
- Nurhanan AR, Wan Rosli WI, Mohsin SSJ (2012). Total polyphenol content and free radical scavenging activity of corn silk (*Zea mays* hairs). *Sains Malays.* 41:1217-1221.
- Pachanawan A, Phumkhachorn P, Rattanachaiyaporn P (2008). Potential of *Psidium guajava* supplemented fish diet in controlling *Aeromonas hydrophila* infection in tilapia (*Oreochromis niloticus*). *J. Biosci. Bioeng.* 106(5):419-424.
- Pradeep K, Raj Mohan C, Gobianand K, Karthikeyan S (2007). Silymarin modulates the oxidant-antioxidant imbalance during diethylnitrosamine induced oxidative stress in rats. *Eur. J. Pharmacol.* 560:110-116.
- Pradhan SC, Girish C (2006). Hepatoprotective herbal drug, silymarin from experimental pharmacology to clinical medicine. *Indian J. Med. Res.* 124:491-504.
- Ren S-H, Liu Z-L, Ding X-L (2009). Isolation and identification of two novel flavones glycosides from corn silk (*Stigma maydis*). *J. Med. Plants Res.* 3:1009-1015.
- Ren S-H, Qiao Q-Q, Ding X-L. (2013). Antioxidative activity of five flavones glycosides from corn silk (*Stigma maydis*). *Czech. J. Food Sci.* 2:148-155.
- Yin G, Ardo L, Thompson KD, Adams A, Jeney Z, Jeney G (2009). Chinese herbs (*Astragalus radix* and *Ganoderma lucidum*) enhance immune response of carp, *Cyprinus carpio*, and protection against *Aeromonas hydrophila*. *Fish Shellfish Immunol.* 26:140-145.
- Zeikoff JT, Wang W, Islam N, Twerdok LE, Curry M, Beaman J, Flesher E (1996). Assays of reactive oxygen intermediates and antioxidant enzymes in medaka (*Oryza latipes*): Potential biomarkers for predicting the effects of environmental pollution. In: *Techniques in Aquatic Toxicology*. (G. Ostrander Ed.), CRC Press, FL., U.S.A., pp. 178-206.



# International Journal of Fisheries and Aquaculture

## Related Journals Published by Academic Journals

- *Journal of Plant Breeding and Crop Science*
- *African Journal of Agricultural Research*
- *Journal of Horticulture and Forestry*
- *International Journal of Livestock Production*
- *International Journal of Fisheries and Aquaculture*
- *Journal of Cereals and Oilseeds*
- *Journal of Soil Science and Environmental Management*
- *Journal of Stored Products and Postharvest Research*

**academicJournals**